Squalene-Derived Triterpene Polyethers from the Red Alga Laurencia omaezakiana

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In our continuing search on halogenated metabolites from species of the red algal genus *Laurencia*, a novel squalene-derived triterpene polyether, named omaezakianol (2), was isolated from *Laurencia omaezakiana* MASUDA along with 15,16-anhydrothyrsiferol (3). Their structures were determined by spectral and chemical methods.

Introduction. – Species of the red algal genus *Laurencia* (Rhodomelaceae, Ceramiales) are known to be prolific sources of a variety of halogenated secondary metabolites, particularly sesquiterpenoids, diterpenoids, triterpenoids, and C_{15} acetogenins [1][2]. Some of these halogenated compounds showed antimicrobial, insecticidal, cytotoxic, and feeding-deterrent activities.

In connection with our chemotaxonomic studies on *Laurencia* species from Japanese waters, we previously reported the isolation and structure elucidation of a pentacyclic triterpene alcohol, enshuol (1), which has a 2,8-dioxabicyclo[5.4.0]undecane molecular skeleton, from *Laurencia omaezakiana* MASUDA ('*Enshu-sozo*' in Japanese), collected at Omaezaki, Shizuoka Prefecture [3]. Although the absolute configurations at C(3), C(6), and C(7) of the 2,8-dioxabicyclo[5.4.0]undecane moiety were determined, the configurations at C(10) to C(22) remained unsettled³). Recently, the structure of enshuol, including the absolute configuration, was established as formula 1 by the asymmetric total synthesis [4]. Further investigation of this species collected at Enoshima, Kanagawa Prefecture, led to the isolation of a novel tetracyclic polyether triterpene alcohol 2, which we named omaezakianol³), along with 15,16-anhydrothyrsiferol³) (3). Herein, we describe the isolation and structure determination of these metabolites.

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³⁾ Trivial atom numbering; for systematic names of 2, 3, and 5-7, see Exper. Part.



Results and Discussion. – A combination of column and thin-layer chromatography of the MeOH extract from the sample collected at Enoshima, Kanagawa Prefecture, gave omaezakianol (2) and 15,16-anhydrothyrsiferol (3) in 15 and 3% yield, respectively, based on the extract.

Omaezakianol (2) was analyzed for $C_{30}H_{52}O_6$ by its HR-FAB-MS. Its IR spectrum showed absorption at $\tilde{\nu}_{max}$ 3455 cm⁻¹ due to OH group(s).

The ¹H- and ¹³C-NMR data (*Table*), together with the ¹H,¹H-COSY, HOHAHA, and HMBC data (*Fig.*) and the FD- and FAB-MS led to the assignment of a planar formula **2**, consisting of four tetrahydrofuran units, for omaezakianol.

The ¹H-NMR spectrum showed the presence of eight tertiary Me groups, five CH H-atoms adjacent to an O-atom, and an olefinic H-atom. As shown in the *Figure*, the ¹H,¹H-COSY and HOHAHA data indicated the presence of a 4-methylpent-3-enyl group, three $-CH_2-CH_2-CH_0-$ moieties, and a $-O-CH-CH_2-CH_2-CH_0-$ moiety. Since **2** has five degrees of unsaturation, **2** must be composed of four O-containing rings. The chemical shift values ($\delta(C)$ 84–86) of the corresponding ring C-atoms in the ¹³C-NMR spectrum suggested that these O-containing rings are tetrahydrofuran moieties. This was confirmed by the mass spectra (FD- and FAB-MS), which showed fragment ions at *m*/*z* 143, 227, and 211 (*Fig.*). The same fragment ions were also observed in the MS of a degradation product **4** derived from enshuol (**1**) by treatment with zinc and AcOH in MeOH [3].

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	$\delta(\mathrm{H})$	$\delta(C)$		$\delta(\mathrm{H})$	$\delta(C)$
Me(1)	1.66 (s)	25.7	CH ₂ (16)	1.70–1.95 (<i>m</i>), 1.95–2.18 (<i>m</i>)	35.8
C(2)	_	131.3	$CH_{2}(17)$	1.38 - 1.60 (m), 1.70 - 1.95 (m)	28.6
H-C(3)	5.04-5.12 (<i>m</i>)	124.8	H - C(18)	4.01 (dd, J = 10.3, 5.9)	84.1
$CH_2(4)$	1.95 - 2.18 (m)	22.4	C(19)	-	85.5
$CH_{2}(5)$	1.25 - 1.36 (m), 1.38 - 1.60 (m)	38.5	$CH_{2}(20)$	1.38 - 1.60 (m), 1.95 - 2.18 (m)	31.0
C(6)	-	73.1	$CH_{2}(21)$	1.70 - 1.95(m), 1.95 - 2.18(m)	26.5
H-C(7)	3.87 (dd, J = 7.1, 7.1)	84.6	H - C(22)	3.82 (dd, J = 7.3, 5.8)	85.5
$CH_{2}(8)$	1.70 - 1.95(m), 1.95 - 2.18(m)	26.2	C(23)	-	71.6
$CH_{2}(9)$	1.38 - 1.60 (m), 1.95 - 2.18 (m)	32.0	Me(24)	1.05 (s)	25.2
C(10)	-	84.8	Me(25)	1.59 (s)	17.6
H - C(11)	3.82 (dd, J = 9.5, 6.1)	85.0	Me(26)	1.21 (s)	24.7
$CH_{2}(12)$	1.38 - 1.60 (m), 1.70 - 1.95 (m)	27.9	Me(27)	1.14 (s)	24.7
$CH_{2}(13)$	1.70 - 1.95(m)	26.5	Me(28)	1.15 (s)	21.0
H - C(14)	3.75 (dd, J = 7.3, 7.3)	83.7	Me(29)	1.13 (s)	25.0
C(15)	_	84.2	Me(30)	1.22(s)	28.1

Table. ¹³C- (DEPT) and ¹H-NMR Data (100 and 400 MHz, resp.; CDCl₃) of Omaezakianol³) (2). Assignments were corroborated by ¹H,¹H-COSY, HSQC, and HMBC experiments. δ in ppm, J in Hz.



m/z 227

Figure. a) Selected fragment ions (m/z) in the FD-MS and FAB-MS and correlations from COSY and HOHAHA; b) long-range correlations from HMBC

by HMBC

Confirmation of the planar structure and determination of the relative configuration of **2** were made as follows. Treatment of omaezakianol (**2**) with 3chloroperbenzoic acid and Na₂HPO₄ in CH₂Cl₂ followed by treatment with a trace of *p*-toluenesulfonic acid gave two isomeric pentacyclic compounds, **5** and **6**, having the molecular formula $C_{30}H_{52}O_7$, in 41 and 44% yield, respectively. Compound **5** showed 15 C-signals in the ¹³C-NMR spectrum, while the isomeric compound **6** showed more than 15 C-signals, thus indicating that **5** and **6** have a symmetrical and unsymmetrical structure, respectively. Fortunately, the symmetrical compound **5** was found to be identical with a synthetic *meso* compound [5], the structure of which was incorrectly proposed for glabrescol that has been isolated from the branches and wood of *Spathelia glabrescens* [6], by comparison of the spectral properties. The weak optical rotation of **5** must be ascribed to a small amount of concomitant impurity. Hence, compound **6** is an epimer at C(3) of **5**. Thus omaezakianol is represented by formula **2** with a $(6R^*,7S^*,10R^*,11S^*,14R^*,15S^*,18R^*,19S^*,22R^*)$ configuration. The asymmetric total synthesis of (+)-omaezakianol is in progress.

Compound **3** having a molecular formula $C_{30}H_{51}BrO_6$, revealed in its ¹H- and ¹³C-NMR spectra signals very similar to those of 15,16-anhydrothyrsiferyl diactate (**7**), which has previously been isolated from *Laurencia saitoi* PERESTENKO (as *Laurencia obtusa* (HUDSON) LAMOUROUX) [7][8]. Extensive spectral analysis by 2D-NMR (¹H,¹H-COSY, HOHAHA, HSQC, HMBC, and NOESY data) strongly suggested **3** to be 15,16-anhydrothyrsiferol (thyrsiferol = $(\alpha^2 S, 2R, 5R) - \alpha^2 - \{(3S) - 3 - \{(2R, 4aR, 6R, 8aS) - 6 - [(2S, 5R) - 5 - bromotetrahydro - 2, 6, 6 - trimethyl - 2H - pyran - 2 - yl]octahydro - 8a - methylpyrano[3, 2 - b]pyran - 2 - yl]-3 - hydroxybutyl}tetrahydro - <math>\alpha^5, \alpha^5, 2$ - trimethylfuran - 2, 5 - dimethanol). Treatment of **3** with Ac₂O and *N*,*N*-dimethylpyridin - 4-amine (DMAP) in CH₂Cl₂ gave the corresponding diacetate, which was identical with 15,16-anhydrothyrsiferyl

diactate (7) in all respects.

To the best of our knowledge, 15,16-anhydrothyrsiferol (**3**) has first been found in the present study. Closely related triterpenoids have previously been isolated from other *Laurencia* species: 15,28-anhydrothyrsiferol (as dehydrothyrsiferol) from *Laurencia pinnatifida* (GMAL. LAMOUR) [9] and 10-epi-15,28- and 10-epi-15,16- anhydrothyrsiferol from *Laurencia viridis* sp. nov [10].

Omaezakianol (2) and 15,16-anhydrothyrsiferol (3) were also isolated from the Omaezaki sample [3] in 7 and 3% yield, respectively; however, enshuol (1) was not found in the Enoshima sample (see *Introduction*). Within the Japanese species of the genus *Laurencia* [2], *L. omaezakiana* was found to produce only triterpenoids as characteristic metabolites. The present study strongly suggests that *L. omaezakiana* may include chemical races as in the case of *Laurencia nipponica* YAMADA [11][12].

Experimental Part

General. Normal-phase high-performance liquid chromatography (HPLC): Shim-Pack Prep-Sil (4.6 mm × 250 mm; Shimadzu). Column chromatography (CC): silica gel (Kieselgel 60, 70–230 mesh; Merck). Prep. TLC: precoated silica gel plate (Kieselgel 60_{2545} , 20 cm × 20 cm, Merck). Optical rotation: Jasco-DIP-140 polarimeter; in CHCl₃. IR Spectra: Jasco-A-102 spectrophotometer; $\tilde{\nu}_{max}$ in cm⁻¹. ¹H- and ¹³C-NMR Spectra: Jeol-JNM-FX-400 spectrometer (at 400 and 100 MHz, resp.); referencing to SiMe₄ as int. standard; δ in ppm, J in Hz. Low- and high-resolution MS: Jeol-JMS-DX303 spectrometer for EI and Jeol-JMS-HX110 spectrometer for FAB and FD; in m/z (rel. intensity).

Plant Material. A specimen of *Laurencia omaezakiana* MASUDA was collected at Enoshima, Kanagawa Prefecture, on 18th August, 1994. A voucher specimen was deposited with the Herbarium of Graduate School of Science, Hokkaido University (SAP 062279).

Extraction and Isolation. Partially dried alga (22.7 g) was immersed in MeOH (300 ml). The MeOH soln. was concentrated, and the residue partitioned between Et_2O and H_2O . The Et_2O soln. was washed with H_2O , dried (Na₂SO₄), and concentrated to give a dark green oil (496 mg). This extract (292 mg) was then fractionated by CC (SiO₂, step gradient hexane/AcOEt). The fraction eluted with AcOEt was further subjected to CC (CHCl₃/MeOH 98:2) followed by prep. TLC (toluene/AcOEt 7:3): 15,16-anhydrothyrsiferol (**3**; 8.8 mg, 3.0% based on the extract). The CC fractions eluted with hexane/AcOEt 3:1 and hexane/AcOEt 1:1 were further subjected to CC (step gradient toluene/AcOEt). The fraction

eluted with toluene/AcOEt 1:1 was then separated by prep. TLC (CHCl₃/MeOH 98:2): omaezakianol (2; 43.8 mg, 15%).

 $\begin{array}{l} Omaezakianol ~(= {\rm rel}-(\alpha^5{\rm R},2{\rm R},2'{\rm S},2''{\rm S},5'{\rm R},5'{\rm R},5''{\rm R},5''{\rm R},5''{\rm R})-Hexadecahydro-\alpha^5,\alpha^{5'''},\alpha^{5''''},\alpha^{5'''},\alpha^{5''''},\alpha^{5'''''''},\alpha^{5'''''''},\alpha^{5'''''''''''''''''''''''$

15,16-Anhydrothyrsiferol (=(α^2 S,2R,5R)- α^2 -{(2E)-3-{(2R,4aR,6R,8aS)-6-{(2S,5R)-5-Bromotetrahydro-2,6,6-trimethyl-2H-pyran-2-yl]octahydro-8a-methylpyrano[3,2-b]pyran-2-yl]but-2-en-1-yl]tetrahy $dro - \alpha^5, \alpha^5, 2$ -trimethylfuran-2,5-dimethanol; **3**). Amorphous solid. $[a]_{2D}^{2D} = +13.1$ (c = 0.53, CHCl₃). IR (CHCl₃): 3650, 3520, 1159, 1147, 1125, 1105, 1078, 1045, 978, 915. ¹H-NMR: 1.13 (s, Me(24)); 1.16 (s, Me(29)); 1.20 (s, Me(26)); 1.21 (br. s, Me(27), Me(30)); 1.27 (s, Me(25)); 1.40 (s, Me(1)); 1.66 (s, Me(28); 1.50-1.93 (m, CH₂(5), CH₂(8), CH₂(9), CH₂(12), H_a-C(13), H_a-C(20), CH₂(21)); 2.10-2.16 $(m, H_a - C(4), H_b - C(13), H_a - C(17), H_b - C(20)); 2.18 - 2.30 (m, H_b - C(4), H_b - C(17)); 3.08 (dd, J = 0.00); 2.18 - 2.30 (m, H_b - C(17), H_b - C(17)); 3.08 (dd, J = 0.00); 2.18 - 2.30 (m, H_b - C(17), H_b - C(17)); 3.08 (dd, J = 0.00); 2.18 - 2.30 (m, H_b - C(17)); 3.08 (dd, J = 0.00); 2.18 - 2.30 (m, H_b - C(17)); 3.08 (dd, J = 0.00); 2.18 - 2.30 (m, H_b - C(17)); 3.08 (dd, J = 0.00); 3.08 ($ 11.5, 2.2, H-C(7); 3.47 (*dd*, J=11.2, 6.4, H-C(11)); 3.56 (*dd*, J=9.8, 2.9, H-C(18)); 3.76 (*dd*, J=12.4); 3.76 (*dd*, J=7.3, 6.8, H-C(16)). ¹³C-NMR: 13.04 (Me(28)); 19.90 (Me(27)); 20.12 (Me(26)); 23.67 (Me(1), Me(29)); 24.03 (Me(24)); 27.71 (Me(30)); 31.04 (Me(25)); 21.82 (CH₂(9)); 23.01 (CH₂(8)); 25.82 (CH₂(13)); 26.57 (CH₂(21)); 28.29 (CH₂(4)); 30.43 (CH₂(17)); 32.01 (CH₂(20)); 37.09 (CH₂(5)); 38.77 (CH₂(12)); 59.05 (CH(3)); 75.20 (CH(14)); 76.72 (CH(18)); 78.38 (CH(11)); 86.63 (CH(7)); 87.60 (CH(22)); 122.3 (CH(16)); 70.48 (C(23)); 72.46 (C(10)); 74.43 (C(6)); 74.69 (C(2)); 85.83 (C(19)); 138.9 (C(15)). FD-MS: 589 and 587 (30 and 32, $[M + H]^+$), 445 and 443 (15 and 17, $[M - C_8H_{15}O_2]^+$), 207 and 205 (20 and 19, $[M - C_{22}H_{37}O_5]^+$), 143 (29, $[M - C_{22}H_{36}BrO_4]^+$). EI-MS: 588 and 586 (0.3 and 0.3, M^+), 570 and 568 (0.5 and 0.5, $[M - H_2O]^+$), 529 and 527 (0.2 and 0.2, $[M - C_3H_7O]^+$), 445 and 443 (0.5 and $0.4, [M - C_8H_{15}O_2]^+), 291 \text{ and } 289 (2.9 \text{ and } 2.8, [M - C_{17}H_{29}O_4]^+), 143 (100, [M - C_{22}H_{36}BrO_4]^+). HR-0.4, [M - C_{17}H_{29}O_4]^+)$ EI-MS: 586.2852 (M^+ , $C_{30}H_{51}^{79}BrO_6^+$; calc. 586.2869).

Transformation of **2** *into* **5** *and* **6**. To a soln. of **2** (10 mg) in CH₂Cl₂ (2 ml) were added 3chloroperbenzoic acid (9 mg) and Na₂HPO₄ · 12 H₂O (5 mg), and the mixture was stirred for 3 h at 0 – 20°. Then a trace of *p*-toluenesulfonic acid was added, and the mixture was further stirred for 1 h. The mixture was extracted with CHCl₃, and the CHCl₃ soln. was successively shaken with sat. Na₂S₂O₃/ Na₂CO₃ soln. 1:2 and sat. brine, dried (Na₂SO₄), and concentrated. The resulting material (13.5 mg) was subjected to HPLC (hexane/CHCl₃/MeOH 625:370:5, flow rate 1 ml/min, RI detector: **5** (4.2 mg, 41%; *t*_R 26 min) and **6** (4.5 mg, 44%; *t*_R 38 min).

[2,2':5',2'':5'',2''':5''',2''''-quinquefuran]-5,5''''-dimethanol (5): Colorless oil. $[\alpha]_{D}^{17} = +0.59$ (c = 1.00, CHCl₃). IR (neat): 3455, 1338, 1319, 1302, 1260, 1183, 1080, 998, 955, 923, 900, 803, 755. ¹H-NMR: 1.06 (s, Me(1), Me(30)); 1.14 (s, Me(26), Me(29)); 1.15 (s, Me(27), Me(28)); 1.24 (s, Me(24), Me(25)); 1.45 (*ddd*, J = 12.1, 8.7, 5.9, $H_a - C(5)$, $H_a - C(20)$); 1.52 - 1.66 (*m*, $H_a - C(8)$, $H_a - C(9)$, $H_a - C(12)$, $H_a-C(13), H_a-C(16), H_a-C(17)); 1.79-1.94$ (m, $H_a-C(4), H_b-C(8), H_b-C(12), H_b-C(13), H_b-C$ $H_{b}-C(17), H_{a}-C(21)); 2.03-2.13 (m, H_{b}-C(4), H_{b}-C(9), H_{b}-C(16), H_{b}-C(21)); 2.25 (ddd, J = 10.13); J = 10.13$ 12.0, 9.3, 7.3, $H_{b}-C(5)$, $H_{b}-C(20)$; 3.73 (br. dd, J=5.4, 5.4, H-C(3), H-C(22)); 3.84 (dd, J=7.8, 5.9, H-C(11), H-C(14)); 4.02 (*dd*, J = 10.3, 5.4, H-C(7), H-C(18)). ¹³C-NMR: 22.57 (Me(27), Me(28)); 25.02 (Me(26), Me(29)); 25.29 (Me(1), Me(30)); 28.09 (Me(24), Me(25)); 26.56 (CH₂(4), CH₂(21)); 27.09 (CH₂(12), CH₂(13)); 29.08 (CH₂(8), CH₂(17)); 31.00 (CH₂(5), CH₂(20)); 34.64 (CH₂(9), CH₂(16)); 84.24 (CH(7), CH(18)); 84.60 (CH(11), CH(14)); 85.68 (CH(3), CH(22)); 71.67 (C(2), C(23); 83.98 (C(10), C(15)); 85.61 (C(6), C(19)). FD-MS: 525 (100, $[M + H]^+$), 381 (25), 227 (29), 143 (27). FAB-MS (pos.): 525 (27, $[M + H]^+$), 507 (20, $[M + H - H_2O]^+$), 391 (23), 227 (31, $[M - H_2O]^+$), 391 (23), 3 $C_{17}H_{29}O_3^{+})$, 154 (100), 143 (54, $[M - C_{22}H_{37}O_4^{+}]^+$). HR-FAB-MS (pos.): 525.3782 ($[M + H]^+$, $C_{30}H_{53}O_7^+$; calc. 525.3792). Spectral data in agreement with those of the synthetic meso compound with a C_s symmetry [5].

rel-(2R,2'S,2''S,2'''S,2'''S,5'R,5''R,5''R,5'''R,5''R,5''R,5''R,5''R,5''R,5''R,5''R,5''R,5''R,5''R,5''R,5''R,5''R,5''R,5''R,5''R,5''R,5'''S,5'''S,5'''S,5'''S,5'''S,5'''S,5''''S,5'''S,5'''S,5'''S,5'

Acetylation of **3** into **7**. To a soln. of **3** (7.0 mg) in CH₂Cl₂ (500 µl) were added Ac₂O (200 µl) and DMAP (10.5 mg), and the mixture was stirred at r.t. for 3 days and then worked up as usual. The product obtained was subjected to TLC (hexane/AcOEt 9:1): $(\alpha^2 S, 2R, 5R) - \alpha^2 - \{(2R, 4aR, 6R, 8aS) - 6[(2S, 5R) - 5-bromotetrahydro-2, 6, 6-trimethyl-2H-pyran-2-yl]octahydro-8a-methylpyrano[3,2-b]pyran-2-yl]but-2-en-1-yl]-tetrahydro-\alpha^5, \alpha^5, 2-trimethylfuran-2, 5-dimethanol diacetate ($ **7** $; 2.2 mg). Amorphous solid. <math>[\alpha]_{D}^{21} = +3.4$ (c = 0.22, CHCl₃). HR-FAB-MS (pos.): 671.3149 ($[M+H]^+$, $C_{34}H_{56}^{29}BrO_8^+$; calc. 671.3159). Further spectral data (¹H- and ¹³C-NMR): in agreement with those of the authentic sample of 15,16-anhydrothyrsiferyl diacetate.

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